

VNU Journal of Science: Natural Sciences and Technology



Journal homepage: https://js.vnu.edu.vn/NST

Original Article

Further Results about Chemical Constituents of *Heterostemma grandiflorum* Cost.

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> Received 26 October 2021 Revised 04 April 2022; Accepted 07 April 2022

Abstract: *Heterostemma grandiflorum* Cost. is a flowering plant of Asclepiadaceae family, distributed in Hoa Binh, Hanoi, Ha Nam,... provinces. We have reported first results about chemical constituents and cytotoxicity of *H. grandiflorum* collected in Me Linh, Hanoi in the previous paper. Further study on dichloromethane and butanol extracts of *H. grandiflorum*'s twigs and leaves, 8 compounds have been isolated. Their structures were elucidated by the 1D and 2D NMR spectroscopy and comparison with published data. They include two flavonoids: apigenin and thalassiolin C; a phenolic: vanillic acid; two triterpenes: 3β -stearyloxy-urs-12-ene, and 3β -acetoxy-22,23,24,25,26,27-hexanordammaran-20-one; and three nitrogen-containing compounds: heteromin A, uracil, and uridine. With exceptions to apigenin and heteromin A, the rest six compounds were found for the first time from *Heterostemma* genus.

Keywords: Heterostemma grandiflorum, flavonoid, triterpene, nitrogen-containing compounds.

1. Introduction

Heterostemma genus belongs to Asclepiadaceae family, consisting of 7 species in Vietnam: H. grandiflorum, H. balansae, H. acuminatum, H. lutea, H. oblongifolium, H. suberosum, and H. villosum. They are

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distributed in many regions such as Hoabinh, Hanoi, Dongnai, Ho Chi Minh City, etc [1]. The traditional medicine of Taiwan used *H. brownii* for cancer treatment and *H. alatum* for sputum consumption and detoxification [2, 3]. According to our literature search, *H. brownii* and *H. alatum* are two species, which were mostly studied for chemical compositions in *Heterostemma* genus [4]. Many compounds of steroids, fatty acids, flavonoids, flavonoid glycosides, adenine, uridine, puriniums and

https://doi.org/10.25073/2588-1140/vnunst.5359

pyrimidines have been isolated from the studied species [2-4]. A series of new compounds: heteromines A - E, I and heteromines F - H have been detected. In particular, heteromines A and B have been reported to show toxic activities against esophageal, liver, lymphadenopathy and leukemia cell lines, while heteromine D has strongly inhibited HL-60 cell lines with the IC₅₀ value of 4.04 nmol/mL [2, 3]. The above mentioned results prompt us to study chemical compositions and cytotoxic activities of H. grandiflorum, collected in Me Linh, Hanoi. The first results about chemical constituents and cytotoxic activities of this species have been published by our group in 2020 [5]. In continuation of our studies on chemical constituents of dichloromethane and n-butanol extracts of H. grandiflorum's leaves and twigs, this paper deals with the isolation and structural elucidation of 8 compounds. Chemical structures were determined by spectroscopic analyses and comparisons with previously published data. They are apigenin (1), thalassiolin C (2), vanillic acid (3), 3β -stearyloxyurs-12-ene (4), 3β-acetoxy-22,23,24,25,26,27hexanordammaran-20-one (5), heteromine A (6), uracil (7) and uridine (8). With exception to apigenin and heteromine A, the rest six compounds (2-5, and 7-8) were isolated as the first presentatives of Heterostemma genus.

2. Experimental

2.1. Plant Materials

Twigs and leaves of *H. grandiflorum* were collected in Me Linh, Ha Noi in October 2019. A voucher specimen (VHH.ML.10.2019.1) is deposited in Institute of Chemistry, VAST, Hanoi, Vietnam. The scientific name was identified by Dr. Nguyen The Cuong, Institute of Ecology and Biological Resources, VAST, Hanoi, Vietnam.

2.2. General Experimental Procedures

NMR: Bruker Avance 500, Germany with TMS as internal reference (for ¹H) and solvent signal (for ¹³C). HR-ESIMS: Agilent 6530 Accurate-Mass Q-TOF LC/MS system.

ESI-MS: 5989B-MS. CC used silica gel 60 G, size 0,043-0,063 mm (Merck), TLC: precoated silica gel G60F₂₅₄ plates (Merck), spots were detected by spraying with vanillin 1% in conc. H₂SO₄ and heating at 110 °C.wigs and leaves of *H. grandiflorum* were collected in Me Linh, Ha Noi in October 2019. A voucher specimen (VHH.ML.10.2019.1) is deposited in Institute of Chemistry, VAST, Hanoi, Vietnam. The scientific name was identified by Dr. Nguyen The Cuong, Institute of Ecology and Biological Resources, VAST, Hanoi, Vietnam.

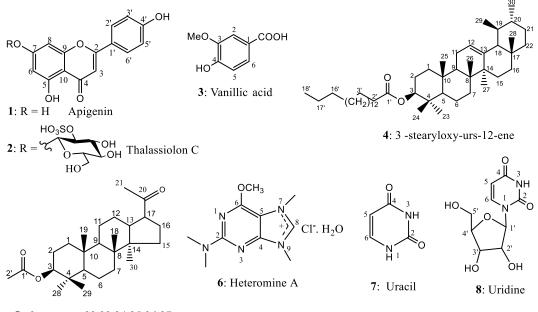
2.3. Extraction and Isolation

The dried leaves and twigs of H. grandiflorum (5,0 kg) were extracted with MeOH:H₂O (95:5) (four times) at room temperature. The methanol extract was concentrated under vacuum and then the partitioned aqueous solution was with *n*-hexane, EtOAc and *n*-BuOH, successively. After solvent evaporation in *vacuum*, *n*-hexane (34,0 g), CH₂Cl₂ (23,0 g) và *n*-BuOH (27,0 g) extracts were obtained.

The dichloromethane extract (23,0 g) was chromatographed on a silica gel column, eluting with gradient *n*-hexane:EtOAc to furnish 7 fractions (D1 - D7). The first one (2,4 g) was rechromatographed on silica gel, eluted with *n*-hexane:CH₂Cl₂ (95:5 - 80:20) to yield 12 subfractions (D1.1 - D1.12). Compound 1 (10 mg), 5 (5 mg) were obtained by purification of subfraction D1.5 (150 mg) using silica gel column, *n*-hexane:CH₂Cl₂ = 90:10. The separation of fraction D3 (2.8 g) was carried out on repeat silica gel columns with suitable eluting solvent systems to furnish 5 mg of compound 4.

The *n*-butanol extract (27,0 g) was given on a Sephadex LH-20 column with MeOH as eluate giving 6 fractions (B1 - B6). Fraction B1 (2.2 g) was reseparated on silica gel column, EtOAc:MeOH (95:5 - 70:30) and then on Sephadex LH-20, MeOH to yield compound **7** (5 mg). Fraction B3 (3,4 g) was purified on RP-18 column, MeOH:H₂O (50:50) to furnish 6 subfractions (B1.1 - B1.6). Repeat chromatography of subfraction B1.3 (350 mg) on Sephadex LH-20 and then on silica gel columns to yield compounds 2 (5 mg) and 3 (4 mg). The separation of fraction B5 (2,9 g) was carried out at first with silica gel column,

n-hexane:acetone (95:5 - 80:20), and then Sephadex LH-20 column, MeOH to yield compounds 6 (7 mg) and 8 (5 mg).



5: 3 -acetoxy-22,23,24,25,26,27hexanordammaran-20-one

Figure 1. Chemical structures of compounds 1-8.

Apigenin (1): (+) ESI-MS: $m/z = 271 [M + H]^+$. ¹H NMR (500 MHz, CD₃OD): $\delta_{\rm H}$ 6.57 (s, H-3), 6.18 (d, J = 2.0 Hz, H-6), 6.41 (d, J = 2.0 Hz, H-8), 7.86 (d, J = 9.0 Hz, H-2', 6'), 6.94 (d, J = 9.0 Hz, H-3', 5').

Thalassiolin C (2): HR-ESIMS: $m/z = 513.0686 \ [M + H]^+$ (calcd. for $C_{21}H_{21}O_{13}S$ 513.0702). ¹H NMR (500 MHz, DMSO-d₆): δ_H 6.868 s (H-3), 6.41 d, 2.5 (H-6), 6.81 d 2.5 (H-8), 7.97 d, 9.0 (H-2', 6'), 6.94 (d 9.0, 1H) (H-3', 5'), 5.28 d, 8.0 (H-1'').

Vanilic acid (3): (-) ESI-MS: m/z = 167 [M - H]^{-. 1}H NMR (500 MHz, CD₃OD): $\delta_{\rm H}$ 7.57 d, 2.0 (H-2), 6.76 d, 8.0 (H-5), 7.48 dd, 2.0 & 8.0 (H-6). ¹³C NMR (125 MHz, CD₃OD): $\delta_{\rm C}$ 129.5 (C-1), 114.1 (C-2), 148.1 (C-3), 150.3 (C-4), 115.2 (C-5), 124.3 (C-6), 173.5 (C-7), 55.7 (3-OCH₃).

 3β -stearyloxy-urs-12-ene (4): (+) ESI-MS: $m/z = 693 [M + H]^+$. ¹H NMR (500 MHz, CDCl₃): $\delta_{\rm H}$ 4.51 (dd, 6.5, 9.0, H-3), 5.13 (t, 3.5, H-12), 1.30 (m, H-18), 1.32 (m, H-20), 0.87 (s, H₃-23), 1.01 (s, H₃-24), 0.98 (s, H₃-25), 0.88 (s, H₃-26), 1.07 (s, H₃-27), 0.80 (s, H₃-28), 0.80 (d, 4.0, H₃-29), 0.89 (d, 4.0, H₃-30), 2.30 (t, 7.5, 14.5, H₂-2'), 0.87 (t, 7.0, H₃-18').

3β-acetoxy-22,23,24,25,26,27 hexanordammaran-20-one (5): (+) ESI-MS: m/z = 403[M + H]⁺. ¹H NMR (500 MHz, CDCl₃): $\delta_{\rm H}$ 4.48 (dd, 5.0, 11.0, H-3), 2.60 (td, 6.0, 11.0, H-17), 0.98 (s, H₃-18), 0.85 (s, H₃-19), 2.13 (s, H₃-21), 0.85 (s, H₃-28), 0.85 (s, H₃-29), 0.87 (s, H₃-30).

Heteromine A (6): HR-ESIMS: m/z = 222.1357 [M]⁺ (calcd. for C₁₀H₁₆N₅O, 222.1354). ¹H NMR (500 MHz, CDCl₃): $\delta_{\rm H}$ 4.20 (s, OMe-6), 4.11 (s, H₃-7), 3.89 (s, H₃-9), 3.29 (s, N-CH₃)₂. ¹³C NMR (125 MHz, CDCl₃): $\delta_{\rm C}$ 162.0 (C-2), 153.5 (C-4), 106.0 (C-5), 159.6 (C-6), 141.0 (C-8), 55.2 (6-OCH₃), 37.7 (N-(CH₃)₂), 36.6 (CH₃-7), 31.4 (CH₃-9).

Uracil (7): HR-ESIMS: m/z = 113.0349[M + H]⁺ (calcd. for C₄H₅N₂O₂ 113.0351). ¹H NMR (500 MHz, CD₃OD): $\delta_{\rm H}$ 5.62 (d, 7.5, H-5), 7.41 (d, 7.5, H-6). ¹³C NMR (125 MHz, CD₃OD): $\delta_{\rm C}$ 153.7 (C-2), 167.4 (C-4), 101.7 (C-5), 143.8 (C-6).

Uridine (8): ¹H-NMR (500 MHz, CD₃OD): $\delta_{\rm H}$ 5.71 (d, 8.0, H-5), 8.01 (d, 8.5, H-6), 5.91 (d, 5.0, H-1'), 4.20 (t, 5.0, H-2'), 4.17 (t, 5.0, H-3'),

4.02 (m, H-4'), 3.85 (dd, 2.5, 12.0, H-5'a), 3.75 (dd, 2.5, 12.0, H-5'b). ¹³C-NMR (125 MHz CD₃OD): 166.3 (C-2), 152.1 (C-4), 90.8 (C-5), 142.7 (C-6), 102.6 (C-1'), 75.7 (C-2'), 71.3 (C-3') 86.4 (C-4'), 62.3 (C-5').

 13 C NMR data of compounds 2, 4 and 5 were given in the Table 1.

Table 1. ¹³C-NMR data (125 MHz) of compounds **2** (DMSO-d₆), 4 (CDCl₃) and 5 (CDCl₃)

	-	-		-			
Position	$\delta_{ m C}$	Position	$\delta_{ m C}$	$\delta_{ m C}$	Position	$\delta_{ m C}$	$\delta_{ m C}$
	2		4	5		4	5
2	164.3	1	38.5 CH ₂	38.8 CH ₂	21	31.3 CH ₂	30.1 CH ₃
3	102.6	2	23.7 CH ₂	23.7 CH ₂	22	41.6 CH ₂	
4	182.0	3	80.6 CH	80.9 CH	23	28.1 CH ₃	
5	161.1	4	37.8 C	37.9 C	24	16.9 CH ₃	
6	99.9	5	55.3 CH	56.0 CH	25	15.7 CH ₃	
7	163.0	6	18.3 CH ₂	18.2 CH ₂	26	16.8 CH ₃	
8	95.1	7	32.9 CH ₂	35.5 CH ₂	27	23.3 CH ₃	
9	157.0	8	40.1 C	40.5 C	28	28.8 CH ₃	28.0 CH ₃
10	105.4	9	47.7 CH	50.7 CH	29	17.5 CH ₃	16.5 CH ₃
1'	120.0	10	36.8 C	37.1 C	30	21.4 CH ₃	15.9 CH ₃
2'	127.7	11	23.4 CH ₂	21.3 CH ₃	1'	173.7 C	171.0 C
3'	116.0	12	124.4 CH	26.0 CH ₂	2'	34.9 CH ₂	21.3 CH ₃
4'	163.0	13	139.7 C	45.2 CH	3'	25.2 CH ₂	
5'	116.0	14	42.1 C	50.1 C	4'-15'	29.7-29.2 CH ₂	
6'	127.7	15	26.7 CH ₂	31.6 CH ₂	16′	31.9 CH ₂	
1″	97.3	16	28.1 CH ₂	25.6 CH ₂	17′	22.7 CH ₂	
2"	78.4	17	33.8 C	54.3 CH	18′	14.1 CH ₃	
3"	75.9	18	59.1 CH	15.6 CH ₃			
4″	69.4	19	39.7 CH	16.3 CH ₃			
5″	76.4	20	39.7 CH	222.4 C			
6"	60.4						

3. Results and Discussion

From dichloromethane and butanol extracts of the twigs and leaves of *H. grandiflorum* eight compounds were isolated by chromatographic methods. The isolates are two flavonoides (1, 2), a phenolic (3), two triterpenes (4, 5) and three nitrogen compounds (6 - 8). Compound 1 was

isolated as light yellow powder. It's ¹H-NMR spectrum indicated typical signals of flavonoid skeleton, including two doublet signals at $\delta_{\rm H}$ 7.86 (d, J = 9.0 Hz, H-2', 6'), 6.94 (d, J = 9.0 Hz, H-3', 5') of ring B and two meta-aromatic protons at $\delta_{\rm H}$ 6.18 (d, J = 2.0 Hz, H-6), 6.41 (d, J = 2.0 Hz, H-8) of ring A and a singlet

signal at $\delta_{\rm H}$ 6.57 (s, H-3) of ring C. These data are total identical with those of apigenin [6]. Many studies have revealed that apigenin has cytotoxic effects on the various cancer cells, atherogenesis, hypertension, prevents the inhibits the asthma, abnormal behavior,... [7]. Compound 2 (yellow powder) was obtained in the mixture with apigenin-7-O- β -Dglucopyranoside (2A), which was reported as compound 3 in our previous study of this plant [5]. Its HR-ESIMS indicated the pseudomolecular peaks at m/z = 433.1134 (C₂₁H₂₁O₁₀) and 513.0686 (C₂₁H₂₁O₁₃S). NMR spectral data of 2 were almost identical with those of apigenin-7-O- β -D-glucopyranoside except only difference on the downfield shift of C-2" ($\Delta\delta_{\rm C} =$ 5.3 ppm). This difference together with 80 mass units more of 2 compared to 2A suggested that this compound has a sulfate group connected at position C-2" of β -D-glucopyranose. Finally, 2 was determined as apigenin-7-O- β -Dglucopyranosyl-2"-sulfate, named thalassiolin C when compared with published literature [7]. Thalassiolins A-C are first flavones having a sulfated β -D-glucose at C-7 position isolated from the Caribbean sea grass Thalassia testudinum [8]. Compound 3 was obtained as a white powder. Its NMR spectra showed signals of a tri-substituted benzene ring revealing by three aromatic protons at $\delta_{\rm H}$ 7.57 d, 2.0 (H-2), 6.76 d, 8.0 (H-5), 7.48 dd, 2.0 & 8.0 (H-6). The substitutes were determined as a methoxy ($\delta_{\rm H}$ 3.88 s, $\delta_{\rm C}$ 55.7), a carboxyl ($\delta_{\rm C}$ 173.5) and a hydroxyl (downfield shift of C-4 at $\delta_{\rm C}$ 150.3) groups. These data led to conclusion that 3 is vanillic acid by comparison with reported data [9].

NMR spectroscopic data of compound 4 (white powder) indicated the typical signals of a derivative of urs-12-ene triterpene, revealed by 6 singlet methyls at $\delta_{\rm H}$ 0.87, 1.01, 0.98, 0.88, 1.07, 0.80 and two doublet methyls at $\delta_{\rm H}$ 0.80 (d, 4.0, H₃-29), 0.89 (d, 4.0, H₃-30); a double bond ($\delta_{\rm H}$ 5.13 t, 3.5, H-12; $\delta_{\rm C}$ 124.4 CH, 139.7 C), an oxy-methine: $\delta_{\rm H}$ 4.51 (dd, 6.5, 9.0, H-3), $\delta_{\rm C}$ 80.6 CH. Beside these signals, the presence of long chain ester group was also observed, including a triplet methyl signal at $\delta_{\rm H}$ 0.87 (t, 7.0, H₃-18'), a methylene connected to carbonyl carbon at $\delta_{\rm H}$ 2.30 (t, 7.0, H₂-2') and a carbonyl ester carbon ($\delta_{\rm C}$ 173.7 C). The connecting position of long chain ester was established at C-3 due to the correlations between C-1' (173.7 ppm) and H-3 (4.51 ppm), H-2' (2.30 ppm) in the HMBC spectrum. ESI-MS of 4 showed a *pseudo-molecular* peak at m/z = 693 [M + H]⁺, corresponding to C₄₈H₈₅O₂. The above-mentioned data were identical with those of 3 β -stearyloxy-urs-12-ene [9]. This compound was isolated previously from the *n*-hexane extract of *Maytenus salicifolia* Reissek (Celastraceae) leaves [10].

¹H and ¹³C-NMR spectra of compound 5 (white powder) indicated the presence of five singlet methyl signals, eight methylenes, five methines with one is oxygenated: $\delta_{\rm H}$ 4.48 dd, 5.0, 11.0; $\delta_{\rm C}$ 80.9 (CH-3), four quaternary carbons, an acetoxyl: $\delta_{\rm H}$ 2.04 s, $\delta_{\rm C}$ 21.3 CH₃, 171.0 C and a methyl-ketone group: $\delta_{\rm H}$ 2.13 s, $\delta_{\rm C}$ 30.1 CH₃, 222.4 C. Comparison of the ¹³C data with those of dammarane-type triterpenes led to conclude that compound 5 is 3β -acetoxyhexanor-dammaran-20-one 22,23,24,25,26,27 and the assignment of ¹³C was given in the table This compound is one of 1 [11, 12]. dammarane-type acetylated triterpenes isolated from methanol extract of Ficus pumila fruit [12].

Compound 6 was separated as white crystals. Its molecular formula C₁₀H₁₆N₅O was deduced from the molecular ion peak at m/z =222.1357 [M]⁺ in the HR-ESIMS spectrum. ¹H and ¹³C NMR data exhibited signals for a dimethylamino group at $\delta_{\rm H}$ 3.29 (s, 6H), $\delta_{\rm C}$ 37.7; two methyl groups connected on two quaternary amines at $\delta_{\rm H}$ 4.11, 3.89; $\delta_{\rm C}$ 36.6, 31.4; one methoxy ($\delta_{\rm H}$ 4.20, $\delta_{\rm C}$ 55.2) and a typical purinium methine CH-8 at $\delta_{\rm H}$ 9.25, $\delta_{\rm C}$ 141.0. Furthermore, the signals of four olefinic quaternary carbons at $\delta_{\rm C}$ 106.0, 153.5, 159.6, 162.0 have been seen. HMBC correlations (3.89/141.0, 153.5; 4.11/141.0, 106.0; 3.29/ 162.0) suggested the structure of 6-methoxy-7,9-dimethyl-2-(N,N dimethylamino) purinium chloride, named heteromine A for 6. This suggestion was confirmed by comparison with those of reported data [13]. Heteromine A together with heteromines B and C were detected as new purinium natural compounds from the aerial parts of *Heterospemma brownii* Hay (Asclepiadaceae), a species used in Taiwan traditional medicine for the treatment of tumors [14]. It was proved to inhibit K562 and HL-60 cell lines [13].

Both compounds 7 and 8 were isolated as yellow powder. Their structures were elucidated as uracil and uridine by comparison with those of published spectral data [15, 16]. Both of these compounds were found in many plant species such as *Colysis hemionitedea, Phlomis samia, Phlomis carica* [4, 16].

4. Conclusion

In this paper we report the further results on chemical constituents of *H. grandiflorum*'s leaves and twigs. In detail, 8 compounds were furnished from dichloromethane and *n*-butanol extracts. Their structures were established as apigenin, thalassiolin C, vanillic acid, 3β -stearyloxy-urs-12-ene, 3β -acetoxy-22,23,24,25, 26,27-hexanordammaran-20-one, heteromine A, uracil and uridine.

Acknowledgements

The research is funded by Vietnam Academy of Science and Technology (VAST) under grant number NCVCC06.10/21-21), and Nguyen Thi Thuy Linh acknowledges financial support by the Ph.D. Scholarship Programme of Vingroup Innovation Foundation (VINIF), code VINIF.2021.TS.071.

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